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Review

Indications for genetic testing in adults with focal segmental glomerulosclerosis



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ABSTRACT

Focal segmental glomerulosclerosis (FSGS) is a histological pattern of injury that derives from various pathological processes that affect podocytes, resulting in loss of selectivity of the glomerular filtration membrane, proteinuria and the development of renal failure that progresses to end-stage kidney disease in a significant number of patients. The classification proposed by the 2021 KDIGO guidelines divides FSGS into four categories: primary, secondary, genetic, and FSGS of undetermined cause, thus facilitating its diagnosis and management. Genetic causes of FSGS present significant clinical variability, complicating their identification. Genetic testing is crucial to identify FSGS of genetic cause. The prevalence of genetic FSGS is significant in children and considerable in adults, highlighting the importance of early diagnosis to avoid unnecessary treatments and facilitate genetic counselling. Massive sequencing techniques have revolutionized genetic diagnosis, allowing the identification of more than 60 genes responsible for podocyte damage. This document proposes clinical recommendations for carrying out genetic studies in adults with FSGS, highlighting the need for a correct classification for adequate therapeutic planning and improvement of results in clinical trials.

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Estudio genético en adultos con glomeruloesclerosis focal y segmentaria

RESUMEN

La glomeruloesclerosis focal y segmentaria (GEFS) es un patrón histológico de lesión que deriva de diversos procesos patológicos que afectan a los podocitos, resultando en pérdida de selectividad del filtrado glomerular, proteinuria y desarrollando insuficiencia renal que progresa a enfermedad renal crónica terminal en un importante número de pacientes. La clasificación propuesta por las guías KDIGO, en 2021, divide la GEFS en cuatro categorías: primaria, secundaria, genética y de causa no determinada, facilitando así su diagnóstico y manejo. Las causas hereditarias de la GEFS presentan una variabilidad clínica significativa, complicando su identificación. El estudio genético es crucial para identificar la GEFS de causa genética. La prevalencia de la GEFS genética es significativa en niños y considerable en adultos, destacando la importancia del diagnóstico temprano para evitar tratamientos innecesarios y facilitar el consejo genético. Las técnicas de secuenciación masiva han revolucionado el diagnóstico genético, permitiendo la identificación de más de 60 genes responsables del daño podocitario. Este documento propone recomendaciones clínicas y patológicas para la realización de estudios genéticos en adultos con GEFS, subrayando la necesidad de una correcta clasificación para la planificación terapéutica adecuada y la mejora de los resultados en ensayos clínicos.

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Palabras clave:

Genética Glomeruloesclerosis focal y segmentaria Indicaciones

Introduction

Focal segmental glomerulosclerosis (FSGS) should be understood as a histologic pattern, not as a disease. This histologic pattern is the result of a variety of pathologic processes that share a common damage on the podocytes; it involves the loss of glomerular filtrate selectivity and clinically it is manifested as proteinuria and long-term renal failure.¹ Histologically, the loss of podocytes involves the migration of epithelial cells from the Bowman's capsule to the glomeruli in an attempt to replace the damaged podocytes, however this attempt of regeneration is inefficient and produces patchy mesangial sclerosis of the glomerular tuft (segmental) that begins in some glomeruli but not all (focal).² The KDIGO (Kidney Disease: Improving Global Outcomes) guidelines for the management of glomerular diseases published in 2021 propose a new etiopathogenic classification, and divided FSGS into four categories, which facilitates the diagnosis and therapeutic approach. Thus there should be different forms: primary, secondary, genetic and of undetermined cause.³ Family and personal history and the clinical presentation are not always sufficient to exclude a hereditary cause of the disease. The clinical presentation of FSGS with a genetic cause is extremely variable; with differences in age of presentation, degree of proteinuria, and progression of chronic kidney disease (CKD).

Initially the genetic forms were described as an onset during childhood, mainly associated with corticosteroid-resistant nephrotic syndrome. However, depending on the selection criteria, up to 30% of adult cases of FSGS may be associated with a genetic cause,^{4–8} so it remains a challenge to define the criteria to perform a genetic study. The clinical and histologic features that appear to better predict a genetic etiology are: the absence of response to immunosuppressive therapies, the presence of microhematuria, the absence of diffuse pedicellar fusion in the renal biopsy, and maintaining a normal serum albumin despite developing nephrotic range proteinuria^{6,9,10} although these last two characteristics may be modified depending on the time of diagnosis.

It is well established that recognizing the diagnosis of genetically caused FSGS is of vital importance for patients. It is clear that, early diagnosis avoids certain diagnostic tests such as scans and exposure to unnecessary immunosuppressive treatment, and also allows the diagnosis of asymptomatic carriers; screening for associated pathologies and facilitates genetic counseling. Unlike the primary forms of FSGS, recurrence of FSGS of genetic cause in renal transplantation is not a problem; but it is relevant in the development of other pathologies such as anti-glomerular basement membrane antibodies which occurs in 2%–3% of renal transplant patients with X-linked Alport syndrome.¹¹ Finally, it also allows an adequate selection of potential living related kidney donors.

A correct classification of patients according to their clinical and histological characteristics is essential for decision making and the planning of a suitable decisions and the planning of an adequate therapeutic scheme. Unfortunately, many clinical trials have failed to because they included patients with different forms of FSGS without a correct stratification.

In the present document we propose recommendations, based on clinical and anatomopathological criteria, for the performance of a genetic study in adult patients with FSGS.

Primary focal and segmental glomerulosclerosis

The etiology of FSGS of immunologic etiology, classically called "primary FSGS"; is not yet fully elucidated. In recent



decades it has been assumed that it is caused by a circulating permeability factor, which is speculated to consist of a group of cytokines that abruptly alter podocyte function, increasing the permeability of the glomerular filtration membrane. This group of cytokines has not yet been defined, and there are several molecules that have been proposed as possible causes of the disease. These include cardiotropic-like cytokine factor-1 (CLCF-1), soluble urokinase-type plasminogen activator receptor (suPAR), the anti-CD40 antibody, apolipoprotein A1 and the soluble form of calcium/calmodulin-serine protein kinase (CASK).¹² In recent years it has been identified that a proportion of patients with minimal change disease are caused by nephrin antibody¹³ and more recently it has been confirmed its role in in primary FSGS especially in relapses after transplant.¹⁴ However, recently it has been found that only 9% of patients with primary FSGS are caused by these antibodies,¹⁵ so it remains to be clarified which are the agents involved in this permeability factor that contributes to the remaining cases of FSGS.

Clinically, primary FSGS is presented abruptly in the form of complete nephrotic syndrome (proteinuria \geq 3.5 g/day, hypoalbuminemia, hypercholesterolemia and edema). Histologically, under the light microscope there are no specific features that would distinguish it from the other other forms of FSGS; by electron microscopy it is highlighted the presence of a diffuse pedicellar fusion that occupies more than 80% of the surface of the glomerular membrane which differentiates it from the secondary forms, and it is a determinant finding. In a series of adult and predominantly white patients with primary FSGS, the median pedicellar fusion was 100%, ¹⁶ however it must be taken into account that several hereditary forms of FSGS can also present with a diffuse pedicellar fusion. A morphometric analysis of pedicle width that excluded patients with hereditary forms of FSGS, found wider pedicels in patients with primary FSGS compared to those with secondary FSGS. A pedicellar width >1500 nm adequately differentiated primary from secondary FSGS.¹⁷ Histologic findings such as podocyte vacuolization and microvillous transformation are related to the amount of proteinuria, and although nonspecific, it is more frequently seen in primary than in secondary forms.¹⁸

The treatment of primary FSGS is immunosuppression, and due to its poor prognosis observed in the absence of treatment, early initiation of immunosuppressive therapy is recommended in cases of FSGS that present with complete nephrotic syndrome.¹⁹

Treatment with glucocorticoids is the current cornerstone of the management of primary FSGS.³ However, this recommendation is based on observational studies and there are no studies that have compared prednisone with placebo in the treatment of primary FSGS.²⁰ A small clinical trial found that the association of mycophenolate with lower doses of steroids achieved similar results with a lower cumulative steroid dose.²¹

The response to corticosteroid therapy is a characteristic feature of primary FSGS, which would rule out other causes of FSGS, although the definition of glucocorticoid resistance is sometimes difficult. Recent studies have even demonstrate a genetic variant in up to 42% of patients with glucocorticoid resistance,²² so that the genetic study would be particularly justified in this group of patients to avoid unnecessary treatment with immunosuppressants.

Calcineurin inhibitors (CNI) are the standard treatment for patients with contraindications, intolerance, or resistance to

Table 1 – Genes associa	ated with focal and	segmental glomerulosc	erosis and their clinical	characteristics.	
Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations
ACTN4 ^{39,41}	19q13.1	Young adult	FSGS, rarely produces NS	AD	None
ANLN ^{39,41}	7p14	Childhood to young adult	FSGS, NSCR	AD	None
APOL1 ^{55,56}	22q12.3	Black individuals	FSGS collapsing collapsing form	AR	Note: there are two risk variants: G1 containing two amino acids (S342G and I384M) near the C-terminus of APOL1. The C terminus of APOL1. G2 is a deletion of two amino acids (del388N389Y) occurring in the same functional domain of APOL1 as G1. These risk haplotypes may be involved in several pathologies in black patients, conferring an increased risk of HIV nephropathy, hypertensive nephropathy, SLE with collapsing features, among others. The bibliography even indicates that this risk is found not only in African or black individuals, but also in individuals from groups with significant recent African ancestry, such as Latinos
ARHGAP24 ^{39,57}	4q21	Adolescence	FSGS	AD	None
ARHGDIA ⁵⁸	17q25.3	Congenital	Congenital nephrotic syndrome	AR	None, the histology may show diffuse mesangial sclerosis (rarely may cause: intellectual deficit, sensorineural deafness, epilepsy and cortical blindness)
CD2AP ^{59,60}	6p12	Childhood, young adult	FSGS, NSCR	AD/AR	None
COL4A3 COL4A4 ^{61,62}	2q36-37, 2q35-57	Young adult the dominant form may manifest itself later in life	FSGS, microhematuria dysmorphic	AD/AR	
COQ8B (previously known as ADCK4) ^{39,63}	19q13	Childhood, adolescence to young adult	NSCR, FSGS	AR	Rare extrarenal manifestations, milder phenotype. CoQ10 deficiecy
EMP2 ³⁹	16p13.2	Childhood	NSCS	AR/AD	Described in families with NSCS
KANK1, KANK2, KANK4 ^{64,65} .	9p24.3, 19p13.2,1p31.3	Few cases reported, <3 years	NSCR	AR	Few cases, some with microhematuria, and some with facial dysmorphia, cardiomyopathy
KIRREL1 ⁶⁶	1q23.1	Childhood until adolescence	NSCR	AR	None
LAMA5 ^{67,68}	20q13.2-q13.3	Childhood to young adulthood	FSGS, NSCR	AD	Lung defects such as bronchial deformity and alveolar dilatation have been described occasionally
LMNA ⁶⁹	1q21.2-q1.3	Young adulthood	FSGS, NSCR	AD	Familial partial lipodystrophy
MAGI2 ⁷⁰	7q11.23-q21.11	Congenital, childhood	Congenital nephrotic syndrome, NSCR FSGS	AR	None
MYO1E (myosin-1E) ⁷¹	15q22.2	Early childhood	NSCR, FSGS	AR	None
MYO9A (myosin 9-A) ⁷²	15q23	Adolescence, young adulthood	FSGS	AD	None, few cases reported
NPHS1 (nephrin) ^{73–75}	19q13.1	Congenital, neonatal, childhood	Congenital nephrotic syndrome	AR	None
NPHS2 (podocin) ^{75–77}	1q25	Congenital, through young adult	Congenital nephrotic syndrome, FSGS, NSCR	AR	None
NUCLEOPORINS:NUP 93, NUP205, NUP85, NUP160, NUP107, NUP133 ^{78–80}	16q13,7q33, 17q25, 11p11.2, 12q15, 1q42.13	Childhood through adolescence	NSCR, FSGS	AR	None. Biopsy may show diffuse mesangial sclerosis NUP93 has been reported to cause collapsing FSGS

– Table 1 (Continued)							
Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations		
PLCE1 ⁸¹	10q23.33	Childhood	NSCR, FSGS	AR	None. Pathology may show diffuse mesangial sclerosis		
PODXL ⁸²	7q32.3	Young adult	FSGS	AD	None		
PTPRO, also known as GLEPP1 ⁸³	12p12.3	Childhood to young adulthood	NSCR FSGS	AR	None		
SGPL1 ^{84,85}	10q22.1	Congenital, neonatal, cases reported in adolescence	NSCR	AR	Few cases reported		
TBC1D8B ^{86,87}	Xq22	May range from neonatal to young adulthood	FSGS, NSCR	LX	None		
TRPC6 ³⁹	11q22	Young adult	FSGS, NSCR	AD	None		
TTC21B ^{88,89}	2q24.3	Childhood to adolescence	FSGS associated with tubulointerstitial lesions	AR/AD	Hypertension and myopia have been reported in some cases		
XPO5 ⁷⁸	6p21.1	Similar to nucleoporins	NSCR, FSGS	AR	None		
ALG1 ^{90,91}	16p13.3	Congenital	Congenital nephrotic syndrome	AR	Associated with glycosylation disorders with microcephaly, developmental delay, abnormal fat distribution, strabismus, and coagulation abnormalities		
AVIL ⁹²	12q14.1	Infancy	NS in the first three years of life, FSGS	AR	It has been associated with microcephaly, short stature, retinal dystrophy, cataracts, deafness, developmental delay		
COL4A5 ^{61,93}	Xq22	Infancy young adult	FSGS, dysmorphic microhematuria	LX	Ocular manifestations: anterior lenticonus, retinopathy (retinal spots), maculopathy. Auditory manifestations: sensorineural hearing loss for high tones		
COQ6 ⁹⁴	14q24.3	Neonatal, infancy some cases up to early adulthood	NSCR, FSGS	AR	Sensorineural deafness. Some patients may present with neurological symptoms such as seizures, encephalopathy, developmental difficulty, occasionally presenting retinopathy. CoQ10 deficiency		
COQ2 ^{63,94}	4q21	Neonatal, childhood. Some cases up to early adulthood	NSCR, FSGS	AR	May present retinopathy, myopathy, multi-organ failure. Neurological and intellectual development symptoms are usually more frequent than in COQ2 and do not present with neurosensory deafness. CoQ10 deficiency		
CRB2 ⁹⁵	9q33	Congenital, neonatal the renal form may present until childhood or adolescence	Congenital nephrotic syndrome, CNRS, FSGS	AR	Some patients present only the renal form, but others may present elevated levels of maternal alpha-fetoprotein, ventriculomegaly/hydrocephalus, in a few patients cardiac and eye defects have been described		
CUBN ⁹⁶	10p13	Childhood	Exceptional FSGS non-nephrotic proteinuria	AR	There have been described cases of isolated FSGS. CUBN has been also associated with another syndrome (when it is homozygous or compound heterozygous); Imerslund-Gräsbeck syndrome: vitamin B12 malabsorption with megaloblastic anemia, growth retardation, recurrent infections, neurological abnormalities, with or without proteinuria, and normal renal function		
EYA1 ¹⁸	8q13.3	Childhood, adult-onset FSGS (when isolated)	FSGS, NSCR, Branchio-oto-renal syndrome	AD	Branchio-oto-renal syndrome, hearing loss auricular malformations, remnants of the branchial arch, and renal abnormalities		

- Table 1 (Continued)						
	Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations
	GLA ^{54,97}	Xq22.1	Youth	FSGS	XL	Fabry disease
	INF2 ⁹⁸	14q32.33	Adolescence, young adult	FSGS Charcot-Marie-Tooth (CMT)	AD	CMT: progressive peripheral sensory-motor neuropathy, muscle weakness, and atrophy, causing inability to walk or grasp objects. Symmetrical amyotrophy, deformity in hands and feet (cavus foot, bowed hands)
	ITGA3 ⁹³	17q21	Congenital	Congenital nephrotic syndrome	AR	Epidermolysis bullosa, interstitial lung disease
	ITGB4, CD151 ^{54,99}	17q25.1, 11p15.5	Congenital	Congenital nephrotic syndrome	AR	Epidermolysis bullosa, pyloric atresia, and occasionally aplasia cutis. They have been associated to congenital NS
	LAMB2 ^{39,93,100}	3p21	Congenital, neonatal, childhood.	NSCR, FSGS, Pierson syndrome	AR	Pierson syndrome: microcoria (extreme nonreactive narrowing of the pupils) due to hypoplasia of the ciliary and pupillary muscles. Many patients die in early childhood, and those who survive often have neurodevelopmental delay and visual loss
	LMX1B ^{101,102}	9q31.1	Infancy to young adulthood	FSGS, nail -patella syndrome	AD	Nail-patella syndrome: presents with hypoplastic or absent patella, dystrophic nails, elbow and iliac horn dysplasia. Open angle glaucoma (about 10% of patients with the syndrome)
	MTTL1, MTTL2, MTTY ^{18,39,103}	MtDNA	Childhood, adult-onset FSGS (when isolated)	FSGS	Mitochondrial	MELAS syndrome: mitochondrial encephalomyopathy, lactic acidosis, stroke episodes. Some cases have been described with isolated FSGS
	MYH9 ¹⁰⁴	22q12	Young adult, sometimes associated with collapsing form	FSGS	AD	Thrombocytopenia with giant platelets, leukocytes with cytoplasmic inclusion bodies (Döhle-like bodies), sensorineural deafness, cataracts at an early age

– Table I (Continuea)					
Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations
					Note: hematological manifestations are usually already present at birth
PAX2 ¹⁰⁵⁻¹⁰⁷	10q24	Adolescent, young adult	FSGS, oculopapillary syndrome	AD	Oculopapillary syndrome presents with CAKUT-type abnormalities associated with extrarenal manifestations such as CNS, ocular and sensorineural hearing loss
PDSS2 ¹⁰⁸	6q21	Congenital, neonatal	Congenital nephrotic syndrome	AR	CoQ10 deficiency, broad neurological phenotype that may present myopathy, seizures, ataxia, developmental difficulties, hypotonia, peripheral neuropathy, sensorineural deafness, etc. Leigh syndrome: growth retardation, ataxia and sensorineural deafness
SCARB2 ^{109,110}	4q21.1	Late childhood or adolescence	NSCR, FSGS, renal-failure action myoclonus syndrome	AR	Renal action-failure myoclonus syndrome: causes progressive myoclonic epilepsy (PME). Renal manifestations often precede neurological symptoms and sometimes only neurological presentation exists without renal involvement. C1q deposits in renal biopsy
SGPL1 ^{84,85}	10q22	Congenital, some cases described up to adolescence	Congenital nephrotic syndrome, NSCR, FSGS	AR	Ichthyosis, adrenal insufficiency, lymphopenia and neurological developmental delay. They can cause diffuse mesangial sclerosis

– Table 1 (Continued)						
Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations	
SMARCAL1 ^{54,95}	2q35	Infancy, but there are mild cases that occur up to adulthood	FSGS, NS	AR	Spondyloepiphyseal dysplasia, T-cell immunodeficiency. 50% of patients may also present hypothyroidism, episodic cerebral ischemia; few patients may have bone marrow insufficiency	
WT1 ^{111,112}	11p13	From childhood to adolescence	FSGS, Frasier syndrome, Denys-Drash syndrome	AD	Frasier syndrome: pseudohermaphroditism in males or ambiguous genitalia, as well as other genital abnormalities such as hypospadias or cryptorchidism, gonad blastoma and rarely they may present nephroblastoma or Wilms tumor, primary amenorrhea. Denys-Drash syndrome: pseudo hermaphroditism in males, gonadal dysgenesis, Wilms tumor, occasionally gonadoblastoma. Primary amenorrhea. In women with XX karyotype there are no abnormalities of the sexual organs, they could only appear as NSCR in adolescence	
WDR73 ⁹³	15q25	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR		
WDR4 ⁹³	21q22	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	Galloway-Mowat syndrome: neurological manifestations (microcephaly, growth retardation, hypotonia, ataxia, behavioral changes). Dysmorphic facial alterations	
TP53RK ⁹³	20q13.12	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	(micrognathia, high-arched palate, hypertelorism, microphthalmos, pointed nose, low-set or drooping ears, anteverted nostrils). Musculoskeletal alterations (scoliosis,	
TPRKB ⁹³	2p13.1	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	pectus excavatum, arachnodactyly, clinodactyly, campylodactyly, dislocated hips, bifid thumb, among others). Cardiac septal defects or dilated cardiomyopathy. Others:	
GON7 ⁹³	14q32.12	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	hypothyroidism, hiatal hernia, hyperpigmented macules, intrauterine growth retardation and oligohydramnios. Note: there are genes in the set that are associated with FSGS as the	
YRDC ⁹³	1p34.3	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	only manifestation, such as the nucleoporin genes	

– Table 1 (Continued)					
Gene	Location	Usual onset	Phenotype	Inheritance	Extra-renal manifestations
NUP107 ⁹³	12q15	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	
NUP133 ⁹³	1q42.13	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	
OSGEP ⁹³	14q11	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	AR	
LAGE3 ⁹³	Xq28	Congenital, neonatal, chilhood	NSCR, FSGS, Galloway-Mowat syndrome	LX	
ZMPSTE24 ¹¹³	1p34.2	Infancy to young adult	FSGS	AR	Mandibuloacral dysplasia: skeletal abnormalities such as hypoplasia of the jaw and clavicles and acro-osteolysis. Kidney disease may be diagnosed later

Other genes that have been associated with FSGS: CLCN5, OCRL, UMOD, NPHP4, DKGE, CFH, EMP2, FAT1, PMM2, DCL1, EXT1, GATA3, MAFB, NXF5, ANKFY1, GAPVD1, MRXA5, CDK20, COG1, E2F3, DHTKD1, SLC35F1.^{54,99,114}

NS: nephrotic syndrome; NSCR: steroid-resistant nephrotic syndrome; NSCS: steroid-sensitive nephrotic syndrome; FSGS: focal segmental glomerulosclerosis; AR: autosomal recessive; AD: autosomal dominant; LX: X-linked; mtDNA: mitochondrial DNA.

Colors: green: FSGS as the only manifestation; orange: FSGS as part of a syndromic picture or with extrarenal manifestations.

glucocorticoids.³ The mechanism of action of glucocorticoids and CNIs in primary FSGS is not entirely clear; presumably these therapies interfere with the cellular sites of the putative permeability factors. However, the information is somewhat ambiguous as to whether the benefit of immunosuppressives depends solely on their immunosuppressive effect or whether it is due to a local effect on podocyte function and the stabilization of synaptopodin.²³ The different series have shown in FSGS a response rate to glucocorticoids and/or CNI of 40%–70% of cases¹⁹; although this proportion is probably underestimated because of the inclusion of patients with secondary or hereditary forms; and probably the rate of response to appropriate immunosuppressive therapy is higher in patients correctly classified as primary FSGS.

Once genetic and secondary causes have been ruled out, the recurrence rate of primary FSGS in renal transplantation is up to 80% in first transplants.²⁴ Early recurrence in transplantation is the hallmark of primary FSGS. The best treatment for recurrent FSGS in transplantation is not yet known. Prophylactic plasmapheresis does not reduce the risk of recurrence, although therapeutic plasmapheresis is considered a good first-line option as a therapy to eliminate permeability factors, especially in very early recurrences with severe proteinuria.²⁵ Prophylactic treatment with rituximab has not been shown to be effective in reducing the risk of recurrence.²⁶

Secondary segmental and focal glomerulosclerosis

Most secondary forms of FSGS are due to a mismatch between glomerular load and glomerular capacity. The pathogenic mechanism is always common and consists of damage and adaptive podocyte loss secondary to increased demand on the glomerular demand (inability to adapt to glomerular size, ischemia due to hypoperfusion). This may occur due to circumstances that reduce renal mass, such as low nephron mass at birth, reflux nephropathy, renal dysplasia, or situations in which there is an increase in the glomerular filtration rate that exceeds the glomerular capacity; as in obesity, high protein intake or androgen abuse.^{27–29} Any chronic glomerular or tubular disease can reduce total nephron function and lead to maladaptive FSGS that overlaps with the primary disorder.³⁰

Hyperfiltration and hypertension in the glomerular capillary represent the main mechanism of strain on the podocyte. Podocytes are extremely sensitive to the "shear stress" generated by increased filtration pressure through the clefts and on their apical surface.^{31,32}

Maladaptive FSGS arises from the processes described above that involve an increase in the glomerular filtration rate of each nephron, leading to a vicious cycle of glomerular hypertrophy, podocyte hypertrophy, podocyte stress and glomerular filtration rate of each nephron, leading to a vicious circle of glomerular hypertrophy, podocyte hypertrophy, podocyte stress and depletion and its depletion with the eventual formation of synechiae and an excess deposition of extracellular matrix within the glomerulus.^{31,33}

Viral infections can induce secondary FSGS, either directly or by release of inflammatory cytokines that interact with podocyte receptors. HIV (human immunodeficiency virus), parvovirus B-19, cytomegalovirus, Epstein–Barr virus, severe acute respiratory syndrome-associated coronavirus-2 (SARS-CoV-2) or simian virus 40 (SV40) are some examples of viral infections that can induce these lesions.^{1,34,35}

There are some drugs that can induce FSGS, such as interferon- α , - β , or - γ , intravenous bisphosphonates, the anthracyclines, lithium, or mTOR (mammalian target of rapamycin) inhibitors. These lesions are usually reversible after discontinuation of treatment.^{28,35}

There is no typical histologic pattern in secondary FSGS, but glomerulomegaly is characteristic of adaptive forms, the perihilar variant (Columbia classification) is also characteristic in this form of FSGS in which synechiae are predominantly present at the perihilar level.³⁵ Collapsing forms are more common incases secondary to HIV or drugs such as bisphosphonates. In contrast, in all forms of secondary FSGS, the pedicellar fusion is patchy usually occupying <40% of the glomerular surface. The presence of diffuse pedicellar fusion exclude the maladaptive mechanism as a cause of primary FSGS.^{18,35}

The biochemical characteristic that differentiates the secondary forms is the presence of normal serum albumin, unlike primary FSGS that debuts with nephrotic syndrome. The temporal presentation of clinical manifestations may also be of great also importance to distinguish between primary and secondary forms. Primary forms usually manifest with an abrupt onset nephrotic syndrome, whereas secondary forms, in particular maladaptive forms, often present with progressive proteinuria. These forms do not respond to immunosuppressive therapy and treatment should be directed, as far as possible, to resolve the underlying cause. Management is based on procedures that decrease glomerular hyperfiltration, withdrawal of the causative drug or treatment of the related infection. Achieving a complete response after treatment with blockade of the renin-angiotensin system (BSRAA), sodiumglucose cotransporter type 2 (iSGLT2) inhibitors³⁶ or treatment of the underlying cause such as weight loss in patients with obesity, would support the diagnosis of secondary FSGS.^{18,37} Recently it has been published, a clinical trial in patients with FSGS in whom treatment with sparsertan (a dual endothelin A and angiotensin II blocker) significantly reduced proteinuria, but without delaying renal progression.³⁸

Although this study was designed to study the effect of sparsentan in patients with presumably primary FSGS, the fact is the median proteinuria of the patients included was in the sub-nephrotic range (urine protein-to-creatinine ratio of 3.1 g/g with a serum albumin within the normal range of (3.49 g/g); indicating that a large proportion of the patients studied did not have primary FSGS but rather secondary or hereditary forms of FSGS, thus indicating its usefulness as an antiproteinuric in the management of these patients.

Segmental and focal glomerulosclerosis secondary to genetic causes

FSGS of genetic or hereditary cause is the least frequent, or perhaps the least diagnosed. In current algorithms, a genetic study is not established as part of the initial workup, so in

Table 2 - Indications for genetic study in adults with focal segmental glomerulosclerosis.

Indications for genetic study in adults with FSGS

- 1. Patients with FSGS diagnosed at any age and family history of CKD
- 2. Patients with FSGS and syndromic symptoms
- 3. Patients with FSGS and family history of consanguinity
- 4. Patients with FSGS diagnosed at any age and presence of persistent microhematuria with dysmorphic red blood cells
- 5. Patients with FSGS and steroid-resistant nephrotic syndrome
- 6. Patients with FSGS and proteinuria of any range with albúmina normal, una vez descartada causa secundaria

this manuscript we want to emphasize that there are adult patients in whom a genetic study would be indicated as a first evaluation before suspicion of FSGS as well as renal biopsy.

Massive sequencing techniques have made it possible to identify more than 60 genes responsible for podocyte damage. The genes encoding proteins located in the podocyte, slit diaphragm, and glomerular basement membrane are the most important involved in genetically caused FSGS, the most frequent being the type IV collagen genes followed by the genes related to podocytopathies (ACTN4, INF2, NPHS1, NPHS2, TRPC6, LMXB1).^{8,39–41} One of the indications for genetic study in adult patients is non-response to immunosuppression, although there are some cases of genetic FSGS that might respond to corticosteroids (EMP2 or PLCG2)³⁹ and some forms alsorespond to CNI (NPHS1, NPHS2, TRPC6, WT1) due to a local effect on the function of podocytes and the stabilization of synaptopodin,²³ inducing a reduction in proteinuria, which can be labeled as partial remission,⁴² but in in genetic forms a complete remission will not be achieved.43,44

The prevalence of genetically caused FSGS is high in children (20%-30%),³⁹ the prevalence in adults ranges from around 22% in patients with a family history of kidney disease to 10% in those with no family history or in undefined cohorts.^{8,40,45,46,41} Hence, genetic testing is generally recommended in patients with early-onset nephrotic syndrome, especially in steroid-resistant forms. However, in the population adult population, the recommendation is more complex. The KDIGO guidelines recommend making an individual assessment of each case to indicate genetic study, considering as characteristics to be taken into account the coexistence of syndromic conditions or family history. The presence of a family history increases the probability that a mutation is responsible for the podocyte damage; although in recently published series, up to 55% of the cases with genetic variants had no evidence of family history.²² This same publication has revealed that in patients with slowly progressive proteinuria without an obvious secondary cause, a genetic variant was found in 33% of the cases studied. This percentage was even higher when the patient had persistent microhematuria with dysmorphic red blood cells, a circumstance that should lead us to suspect a disease related to collagen IV.

Electron microscopy is not useful for diagnosing or suspecting FSGS of genetic etiology. Those forms with complete nephrotic syndrome at the time of renal biopsy will show extensive pedicellar fusion while those without nephrotic syndrome will show only partial pedicellar fusion. Occasionally, in FSGS associated with collagen IV mutations the electron microscopy shows a thin glomerular basement membrane (<180 nm in children and <250 nm in adults),⁴⁷ which could be an indication for genetic study.

Currently the most widely used method for genetic diagnosis is massive sequencing. These techniques allow the simultaneous sequencing of the exons of a set of genes, or of all the exons (exome sequencing) or of the entire genome. The Sanger technique is currently used to sequence small genes or to confirm a variant previously identified by mass sequencing.^{48–50} Gene panels associated with various diseases have proven to be useful in nephrology, so they are a good tool when we are looking for diseases without a specific phenotype as in the case of FSGS of genetic cause.^{51,52}

Indications for genetic study in GEFS

In accordance with the situations of routine clinical practice and the difficulties that may arise in certain centers regarding the access to genetic studies, we have proposed the following algorithm: (Fig. 1).

Although many of the genes that cause FSGS give rise to a disease with onset in childhood, the wide phenotypic spectrum does not allow us to rule out these genes as the cause of adult-onset disease. For this reason, gene panels should include all genes causing glomerular nephropathy.

Some genes that are the cause for other inherited diseases such as tubulopathies (CLCN5, OCRL), thrombotic microangiopathy (DKGE, CFH), ciliopathies (UMOD, NPHP4),⁵³ deposit disease such as Fabry disease (GLA)³⁹ and CAKUT-causing genes may present a FSGS pattern in the biopsy.⁵⁴

Table 1 describes the genes that have been associated with FSGS of genetic cause and their clinical features. They have been divided according to whether patients present with extrarenal manifestations as part of a syndromic picture or whether they are the cause of FSGS as the only clinical expression. To date there are more than 60 associated genes; however, we have to take into account that as research progresses new genes are being discovered and FSGS may be due to alterations of the podocyte, the glomerular basement membrane and its components, but it may also be the scar of renal disease of any origin⁵⁴ (Table 1).

In Table 2 we summarize the indications for genetic study in patients with FSGS.

Declaration of competing interest

Nothing declared.

$\mathbf{R} \to \mathbf{F} \to \mathbf{R} \to \mathbf{N} \to \mathbf{C} \to \mathbf{S}$

- Shabaka A, Tato Ribera A, Fernández-Juárez G. Focal segmental glomerulosclerosis: state-of-the-art and clinical perspective. Nephron. 2020;144:413–27, http://dx.doi.org/10.1159/000508099.
- Rosenberg AZ, Kopp JB. Focal segmental glomerulosclerosis. Clin J Am Soc Nephrol. 2017;12:502–17, http://dx.doi.org/10.2215/CJN.05960616.
- Kidney Disease: Improving Global Outcomes (KDIGO) Glomerular Diseases Work Group. KDIGO 2021 clinical practice guideline for the management of glomerular diseases. Kidney Int. 2021;100:S1–276, http://dx.doi.org/10.1016/j.kint.2021.05.021.
- Gribouval O, Boyer O, Hummel A, Dantal J, Martinez F, Sberro-Soussan R, et al. Identification of genetic causes for sporadic steroid-resistant nephrotic syndrome in adults. Kidney Int. 2018;94:1013–22,

http://dx.doi.org/10.1016/j.kint.2018.07.024.

- Gast C, Pengelly RJ, Lyon M, Bunyan DJ, Seaby EG, Graham N, et al. Collagen (COL4A) mutations are the most frequent mutations underlying adult focal segmental glomerulosclerosis. Nephrol Dial Transplant. 2016;31:961–70, http://dx.doi.org/10.1093/ndt/gfv325.
- Miao J, Pinto e Vairo F, Hogan MC, Erickson SB, El Terds M, Bentall AJ, et al. Identification of genetic causes of focal segmental glomerulosclerosis increases with proper patient selection. Mayo Clin Proc. 2021;96:2342–53, http://dx.doi.org/10.1016/j.mayocp.2021.01.037.
- Braunisch MC, Riedhammer KM, Herr PM, Draut S, Günthner R, Wagner M, et al. Identification of disease-causing variants by comprehensive genetic testing with exome sequencing in adults with suspicion of hereditary FSGS. Eur J Hum Genet. 2021;29:262–70,

http://dx.doi.org/10.1038/s41431-020-00719-3.

- Yao T, Udwan K, John R, Rana A, Haghighi A, Xu L, et al. Integration of genetic testing and pathology for the diagnosis of adults with FSGS. Clin J Am Soc Nephrol. 2019;14:213–23, http://dx.doi.org/10.2215/CJN.08750718.
- Santín S, Bullich G, Tazón-Vega B, García-Maset R, Giménez I, Silva I, et al. Clinical utility of genetic testing in children and adults with steroid-resistant nephrotic syndrome. Clin J Am Soc Nephrol. 2011;6:1139–48, http://dx.doi.org/10.2215/CIN.05260610

http://dx.doi.org/10.2215/CJN.05260610.

- Lepori N, Zand L, Sethi S, Fernandez-Juarez G, Fervenza FC. Clinical and pathological phenotype of genetic causes of focal segmental glomerulosclerosis in adults. Clin Kidney J. 2018;11:179–90, http://dx.doi.org/10.1093/ckj/sfx143.
- Savige J, Lipska-Zietkiewicz BS, Watson E, Hertz JM, Celtas C, Mari F, et al. Guidelines for genetic testing and management of Alport syndrome. Clin J Am Soc Nephrol CJASN. 2022;17:143–54, http://dx.doi.org/10.2215/CJN.04230321.
- Königshausen E, Sellin L. Circulating permeability factors in primary focal segmental glomerulosclerosis: a review of proposed candidates. BioMed Res Int. 2016;2016:1–9, http://dx.doi.org/10.1155/2016/3765608.
- Watts AJB, Keller KH, Lerner G, Rosales I, Collins AB, Sekulic M, et al. Discovery of autoantibodies targeting nephrin in minimal change disease supports a novel autoimmune etiology. J Am Soc Nephrol JASN. 2022;33:238–52, http://dx.doi.org/10.1681/ASN.2021060794.
- 14. Shirai Y, Miura K, Ishizuka K, Ando T, Kanda S, Hashimoto J, et al. A multi-institutional study found a possible role of anti-nephrin antibodies in post-transplant focal segmental glomerulosclerosis recurrence. Kidney Int. 2024;105:608–17, http://dx.doi.org/10.1016/j.kint.2023.11.022.

- Hengel FE, Dehde S, Lassé M, Zahner G, Seifert L, Schnarre A, et al. Autoantibodies targeting nephrin in podocytopathies. N Engl J Med. 2024;391:422–33, http://dx.doi.org/10.1056/NEJMoa2314471.
- Hommos MS, De Vriese AS, Alexander MP, Sethi S, Vaughan L, Zand L, et al. The incidence of primary vs secondary focal segmental glomerulosclerosis: a clinicopathologic study. Mayo Clin Proc. 2017;92:1772–81, http://dx.doi.org/10.1016/j.mayocp.2017.09.011.
- Deegens JKJ, Dijkman HBPM, Borm GF, Steenbergen EJ, van den Berg JG, Weening JJ, et al. Podocyte foot process effacement as a diagnostic tool in focal segmental glomerulosclerosis. Kidney Int. 2008;74:1568–76, http://dx.doi.org/10.1038/ki.2008.413.
- De Vriese AS, Sethi S, Nath KA, Glassock RJ, Fervenza FC. Differentiating primary, genetic, and secondary FSGS in adults: a clinicopathologic approach. J Am Soc Nephrol. 2018;29:759–74, http://dx.doi.org/10.1681/ASN.2017090958.
- Korbet SM. Treatment of primary FSGS in adults. J Am Soc Nephrol. 2012;23:1769–76, http://dx.doi.org/10.1681/ASN.2012040389.
- 20. Hodson EM, Sinha A, Cooper TE. Interventions for focal segmental glomerulosclerosis in adults. Cochrane Database Syst Rev. 2022;2,

http://dx.doi.org/10.1002/14651858.CD003233.pub3.

- Senthil Nayagam L, Ganguli A, Rathi M, Kohli HS, Gupta KL, Joshi K, et al. Mycophenolate mofetil or standard therapy for membranous nephropathy and focal segmental glomerulosclerosis: a pilot study. Nephrol Dial Transplant. 2008;23:1926–30, http://dx.doi.org/10.1093/ndt/gfm538.
- Tato AM, Carrera N, García-Murias M, Shabaka A, Ávila A, Mora Mora MT, et al. Genetic testing in focal segmental glomerulosclerosis: in whom and when? Clin Kidney J. 2023;16:2011–22, http://dx.doi.org/10.1093/ckj/sfad193.
- Faul C, Donnelly M, Merscher-Gomez S, Chang YH, Franz S, Delfgaauw J, et al. The actin cytoskeleton of kidney podocytes is a direct target of the antiproteinuric effect of cyclosporine A. Nat Med. 2008;14:931–8, http://dx.doi.org/10.1038/nm.1857.
- 24. Maas RJH, Deegens JKJ, van den Brand JAJG, Cornelissen EAM, Wetzels JFM. A retrospective study of focal segmental glomerulosclerosis: clinical criteria can identify patients at high risk for recurrent disease after first renal transplantation. BMC Nephrol. 2013;14:47, http://dx.doi.org/10.1186/1471-2369-14-47.
- Campise M, Favi E, Messa P. Clinical outcomes of prophylactic and therapeutic plasmapheresis in adult deceased-donor kidney transplant recipients with primary focal segmental glomerulosclerosis. Exp Clin Transplant. 2019;17, http://dx.doi.org/10.6002/ect.2018.0106.
- 26. Auñón P, Polanco N, Pérez-Sáez MJ, Rodrigo E, Sancho A, Pascual J, et al. Pre-emptive rituximab in focal and segmental glomerulosclerosis patients at risk of recurrence after kidney transplantation. Clin Kidney J. 2021;14:139–48, http://dx.doi.org/10.1093/ckj/sfz120.
- Rosenberg AZ, Kopp JB. Focal segmental glomerulosclerosis. Clin J Am Soc Nephrol. 2017;12:502–17, http://dx.doi.org/10.2215/CJN.05960616.
- Fogo AB. Causes and pathogenesis of focal segmental glomerulosclerosis. Nat Rev Nephrol. 2015;11:76–87, http://dx.doi.org/10.1038/nrneph.2014.216.
- D'Agati VD, Chagnac A, De Vries APJ, Levi M, Porrini E, Herman-Edelstein M, et al. Obesity-related glomerulopathy: clinical and pathologic characteristics and pathogenesis. Nat Rev Nephrol. 2016;12:453–71, http://dx.doi.org/10.1038/nrneph.2016.75.
- 30. Rennke HG, Klein PS. Pathogenesis and significance of nonprimary focal and segmental glomerulosclerosis. Am J

Kidney Dis. 1989;13:443–56, http://dx.doi.org/10.1016/S0272-6386(89)80001-0.

- Kriz W, Lemley KV. A potential role for mechanical forces in the detachment of podocytes and the progression of CKD. J Am Soc Nephrol. 2015;26:258–69, http://dx.doi.org/10.1681/ASN.2014030278.
- Kriz W, Lemley KV. Potential relevance of shear stress for slit diaphragm and podocyte function. Kidney Int. 2017;91:1283–6, http://dx.doi.org/10.1016/j.kint.2017.02.032.
- Cortinovis M, Perico N, Ruggenenti P, Remuzzi A, Remuzzi G. Glomerular hyperfiltration. Nat Rev Nephrol. 2022;18:435–51, http://dx.doi.org/10.1038/s41581-022-00559-y.
- 34. Kudose S, Batal I, Santoriello D, Xu K, Barasch J, Peleg Y, et al. Kidney biopsy findings in patients with COVID-19. J Am Soc Nephrol. 2020;31:1959–68, http://dx.doi.org/10.1681/ASN.2020060802.
- D'Agati VD, Kaskel FJ, Falk RJ. Focal segmental glomerulosclerosis. N Engl J Med. 2011;365:2398–411, http://dx.doi.org/10.1056/NEJMra1106556.
- 36. Yau K, Kuah R, Cherney DZI, Lam TKT. Obesity and the kidney: mechanistic links and therapeutic advances. Nat Rev Endocrinol. 2024;20:321–35, http://dx.doi.org/10.1038/s41574-024-00951-7.
- Praga M, Borstein B, Andres A, Arenas J, Oliet A, Montoyo C, et al. Nephrotic proteinuria without hypoalbuminemia: clinical characteristics and response to angiotensin-converting enzyme inhibition. Am J Kidney Dis. 1991;17:330–8,

http://dx.doi.org/10.1016/s0272-6386(12)80483-5.

- Rheault MN, Alpers CE, Barratt J, Bieler S, Canetta P, Chae DW, et al. Sparsentan versus irbesartan in focal segmental glomerulosclerosis. N Engl J Med. 2023;389:2436–45, http://dx.doi.org/10.1056/NEJMoa2308550.
- Sambharia M, Rastogi P, Thomas CP. Monogenic focal segmental glomerulosclerosis: a conceptual framework for identification and management of a heterogeneous disease. Am J Med Genet C Semin Med Genet. 2022;190:377–98, http://dx.doi.org/10.1002/AJMG.C.31990.
- 40. Gast C, Pengelly RJ, Lyon M, Bunyan DJ, Seaby EG, Graham N, et al. Collagen (COL4A) mutations are the most frequent mutations underlying adult focal segmental glomerulosclerosis. Nephrol Dial Transplant. 2016;31:961–70, http://dx.doi.org/10.1093/ndt/gfv325.
- Sen ES, Dean P, Yarram-Smith L, Bierzynska A, Woodward G, Buxton C, et al. Clinical genetic testing using a custom-designed steroid-resistant nephrotic syndrome gene panel: analysis and recommendations. J Med Genet. 2017;54:795–804,

http://dx.doi.org/10.1136/jmedgenet-2017-104811.

- 42. Li R, Dong W, Chen Y, Tang T, Zhao X, Zhang L, et al. Effect of cyclosporine A on focal segmental glomerulosclerosis caused by MYO1E mutation in a Chinese adult patient: a case report. Medicine (Baltimore). 2023;102:e32683, http://dx.doi.org/10.1097/MD.00000000032683.
- Gubler MC. Nephrotic syndrome: genetic testing in steroid-resistant nephrotic syndrome. Nat Rev Nephrol. 2011;7:430–1, http://dx.doi.org/10.1038/nrneph.2011.75.
- De Vriese AS, Sethi S, Nath KA, Glassock RJ, Fervenza FC. Differentiating primary, genetic, and secondary FSGS in adults: a clinicopathologic approach. J Am Soc Nephrol. 2018;29:759–74, http://dx.doi.org/10.1681/ASN.2017090958.
- 45. Connaughton DM, Kennedy C, Shril S, Mann N, Murray SL, Williams PA, et al. Monogenic causes of chronic kidney disease in adults. Kidney Int. 2019;95:914–28, http://dx.doi.org/10.1016/j.kint.2018.10.031.
- 46. Groopman EE, Marasa M, Cameron-Christie S, Petrovski S, Aggarwal VS, Milo-Rasouly H, et al. Diagnostic utility of

exome sequencing for kidney disease. N Engl J Med. 2019;380:142–51, http://dx.doi.org/10.1056/nejmoa1806891.

- Uzzo M, Moroni G, Ponticelli C. Thin basement membrane: an underrated cause of end-stage renal disease. Nephron. 2023;147:383–91, http://dx.doi.org/10.1159/000528243.
- 48. Knoers N, Antignac C, Bergmann C, Dahan K, Giglio S, Heidet L, et al. Genetic testing in the diagnosis of chronic kidney disease: recommendations for clinical practice. Nephrol Dial Transplant. 2022;37:239–54, http://dx.doi.org/10.1093/ndt/gfab218.
- 49. Ars E. Enfoque genético de las enfermedades renales hereditarias. In: Lorenzo V, Lopez Gomez J.M. (Eds). Nefrología al día [Accessed 15 June 2024]. Available from: https://nefrologiaaldia.org/es-articulo-enfoque-geneticoenfermedades-renales-hereditarias-359.
- Groopman EE, Rasouly HM, Gharavi AG. Genomic medicine for kidney disease. Nat Rev Nephrol. 2018;14:83–104, http://dx.doi.org/10.1038/nrneph.2017.167.
- Domingo-Gallego A, Pybus M, Bullich G, Furlano M, Ejarque-Vila L, Lorente-Grandoso L, et al. Clinical utility of genetic testing in early-onset kidney disease: seven genes are the main players. Nephrol Dial Transplant. 2021;37:687–96, http://dx.doi.org/10.1093/ndt/gfab019.
- 52. Bullich G, Domingo-Gallego A, Vargas I, Ruiz P, Lorente-Grandoso L, Furlano M, et al. A kidney-disease gene panel allows a comprehensive genetic diagnosis of cystic and glomerular inherited kidney diseases. Kidney Int. 2018;94:363–71, http://dx.doi.org/10.1016/j.kint.2018.02.027.
- 53. Chun J, Wang M, Wilkins MS, Knob AU, Benjamin A, Bu L, et al. Autosomal dominant tubulointerstitial kidney disease—uromodulin misclassified as focal segmental glomerulosclerosis or hereditary glomerular disease. Kidney Int Rep. 2020;5:519–29,
 - http://dx.doi.org/10.1016/j.ekir.2019.12.016.
- 54. Mitrotti A, Giliberti M, Di Leo V, Di Bari I, Pontrelli P, Gesualdo L. Hidden genetics behind glomerular scars: an opportunity to understand the heterogeneity of focal segmental glomerulosclerosis? Pediatr Nephrol. 2024;39:1685–707, http://dx.doi.org/10.1007/s00467-023-06046-1.
- Friedman DJ, Pollak MR. Apol1 nephropathy: from genetics to clinical applications. Clin J Am Soc Nephrol. 2021;16:294–303, http://dx.doi.org/10.2215/CJN.15161219.
- Friedman DJ, Pollak MR. APOL1 and kidney disease: from genetics to biology. Annu Rev Physiol. 2020;82:323–42, http://dx.doi.org/10.1146/annurev-physiol-021119-034345.
- 57. Gbadegesin RA, Winn MP, Smoyer WE. Genetic testing in nephrotic syndrome — challenges and opportunities. Nat Rev Nephrol. 2013;9:179–84, http://dx.doi.org/10.1038/nrneph.2012.286.
- 58. Gupta IR, Baldwin C, Auguste D, Ha KC, El Andalousi J, Fahiminiya S, et al. ARHGDIA: a novel gene implicated in nephrotic syndrome. J Med Genet. 2013;50:330–8, http://dx.doi.org/10.1136/jmedgenet-2012-101442.
- 59. Gigante M, Pontrelli P, Montemurno E, Roca L, Aucella F, Penza R, et al. CD2AP mutations are associated with sporadic nephrotic syndrome and focal segmental glomerulosclerosis (FSGS). Nephrol Dial Transplant. 2009;24:1858–64, http://dx.doi.org/10.1093/ndt/gfn712.
- 60. Takano T, Bareke E, Takeda N, Aoudjit L, Baldwin C, Pisano P, et al. Recessive mutation in CD2AP causes focal segmental glomerulosclerosis in humans and mice. Kidney Int. 2019;95:57–61, http://dx.doi.org/10.1016/j.kint.2018.08.014.
- Savige J, Harraka P. Pathogenic variants in the genes affected in Alport syndrome (COL4A3–COL4A5) and their association with other kidney conditions: a review. Am J Kidney Dis. 2021;78:857–64, http://dx.doi.org/10.1053/j.ajkd.2021.04.017.
- 62. Solanki KV, Hu Y, Moore BS, Abedi V, Avula V, Mirshahi T, et al. The phenotypic spectrum of COL4A3 heterozygotes.

Kidney Int Rep. 2023;8:2088–99, http://dx.doi.org/10.1016/j.ekir.2023.07.010.

63. Drovandi S, Lipska-Ziętkiewicz BS, Ozaltin F, Emma F, Gulhan B, Boyer O, et al. Variation of the clinical spectrum and genotype-phenotype associations in Coenzyme Q10 deficiency associated glomerulopathy. Kidney Int. 2022;102:592–603, http://dx.doi.org/10.1016/j.kipt.2022.02.040

http://dx.doi.org/10.1016/j.kint.2022.02.040.

- Allison SJ. The KANK family in podocyte function. Nat Rev Nephrol. 2015;11:79504, http://dx.doi.org/10.1038/nrneph.2015.84.
- Gee HY, Zhang F, Ashraf S, Kohl S, Sadowski CE, Vega-Warner V, et al. KANK deficiency leads to podocyte dysfunction and nephrotic syndrome. J Clin Invest. 2015;125:2375–84, http://dx.doi.org/10.1172/JCI79504.
- 66. Solanki AK, Widmeier E, Arif E, Sharma S, Daga A, Srivastava P, et al. Mutations in KIRREL1, a slit diaphragm component, cause steroid-resistant nephrotic syndrome. Kidney Int. 2019;96:883–9, http://dx.doi.org/10.1016/j.kint.2019.06.016.
- 67. Kaimori JY, Kikkawa Y, Motooka D, Namba-Hamano T, Takuwa A, Okazaki A, et al. A heterozygous LAMA5 variant may contribute to slowly progressive, vinculin-enhanced familial FSGS and pulmonary defects. JCI Insight. 2022;7(23), http://dx.doi.org/10.1172/jci.insight.158378.
- Savige J, Harraka P. Pathogenic LAMA5 variants and kidney disease. Kidney360. 2021;2:1876–9, http://dx.doi.org/10.34067/kid.0007312021.
- 69. Thong KM, Xu Y, Cook J, Takou A, Wagner B, Kawar B, et al. Cosegregation of focal segmental glomerulosclerosis in a family with familial partial lipodystrophy due to a mutation in LMNA. Nephron Clin Pract. 2013;124(1-2):31-7, http://dx.doi.org/10.1159/000354716.
- Bierzynska A, Soderquest K, Dean P, Colby E, Rollason R, Jones C, et al. MAGI2 mutations cause congenital nephrotic syndrome. J Am Soc Nephrol. 2017;28:1614–21, http://dx.doi.org/10.1681/ASN.2016040387.
- 71. Krendel M, Leh S, Garone ME, Edwards-Richards A, Lin JJ, Brackman D, et al. Focal segmental glomerulosclerosis and proteinuria associated with Myo1E mutations: novel variants and histological phenotype analysis. Pediatr Nephrol. 2023;38:439–49, http://dx.doi.org/10.1007/s00467-022-05634-x.
- 72. Li Q, Gulati A, Lemaire M, Nottoli T, Bale A, Tufro A. Rho-GTPase Activating Protein myosin MYO9A identified as a novel candidate gene for monogenic focal segmental glomerulosclerosis. Kidney Int. 2021;99:1102–17, http://dx.doi.org/10.1016/j.kint.2020.12.022.
- Kaukinen A, Kuusniemi AM, Lautenschlager I, Jalanko H. Glomerular endothelium in kidneys with congenital nephrotic syndrome of the Finnish type (NPHS1). Nephrol Dial Transplant. 2008;23:1224–32, http://dx.doi.org/10.1093/ndt/gfm799.
- 74. Patrakka J, Kestilä M, Wartiovaara J, Ruotsalainen V, Tissari P, Lenkkeri U, et al. Congenital nephrotic syndrome (NPHS1): features resulting from different mutations in Finnish patients. Kidney Int. 2000;58:972–80, http://dx.doi.org/10.1046/j.1523-1755.2000.00254.x.
- Pollak MR. The genetic basis of FSGS and steroid-resistant nephrosis. Semin Nephrol. 2003;23:141–6, http://dx.doi.org/10.1053/SNEP.2003.50014.
- 76. Tonna SJ, Needham A, Polu K, Uscinski A, Appel GB, Falk RJ, et al. NPHS2 variation in focal and segmental glomerulosclerosis. BMC Nephrol. 2008;9:13, http://dx.doi.org/10.1186/1471-2369-9-13.
- Sadowski CE, Lovric S, Ashraf S, Pabst WL, Gee HY, Kohl S, et al. A single-gene cause in 29.5% of cases of steroid-resistant nephrotic syndrome. J Am Soc Nephrol. 2015;26:1279–89, http://dx.doi.org/10.1681/ASN.2014050489.

- Braun DA, Sadowski CE, Kohl S, Lovric S, Astrinidis SA, Pabst WL, et al. Mutations in nuclear pore genes NUP93, NUP205 and XPO5 cause steroid-resistant nephrotic syndrome. Nat Genet. 2016;48:457–65, http://dx.doi.org/10.1038/ng.3512.
- Braun DA, Lovric S, Schapiro D, Schneider R, Marquez J, Asif M, et al. Mutations in multiple components of the nuclear pore complex cause nephrotic syndrome. J Clin Invest. 2018;128:4313–28, http://dx.doi.org/10.1172/JCI98688.
- 80. Cason RK, Williams A, Chryst-Stangl M, Wu G, Huggins K, Brathwaite KE, et al. Collapsing focal segmental glomerulosclerosis in siblings with compound heterozygous variants in NUP93 expand the spectrum of kidney phenotypes associated with nucleoporin gene mutations. Front Pediatr. 2022;10:1–7, http://dx.doi.org/10.3389/fped.2022.915174.
- Hinkes B, Wiggins RC, Gbadegesin R, Vlangos CN, Seelow D, Nürnberg G, et al. Positional cloning uncovers mutations in PLCE1 responsible for a nephrotic syndrome variant that may be reversible. Nat Genet. 2006;38:1397–405, http://dx.doi.org/10.1038/ng1918.
- Marx D, Caillard S, Olagne J, Moulin B, Hannedouche T, Touchard G, et al. Atypical focal segmental glomerulosclerosis associated with a new PODXL nonsense variant. Mol Genet Genomic Med. 2021;9:1–9, http://dx.doi.org/10.1002/mgg3.1658.
- Ozaltin F, Ibsirlioglu T, Taskiran EZ, Baydar DE, Kaymaz F, Buyukcelik M, et al. Disruption of PTPRO causes childhood-onset nephrotic syndrome. Am J Hum Genet. 2011;89:139–47, http://dx.doi.org/10.1016/j.ajhg.2011.05.026.
- 84. Carney EF. SGPL1 mutations cause a novel SRNS syndrome. Nat Rev Nephrol. 2017;13:191,
- http://dx.doi.org/10.1038/nrneph.2017.19.
 85. Tastemel Ozturk T, Canpolat N, Saygili S, Bayrakci US, Soylemezoglu O, Ozaltin F, et al. A rare cause of nephrotic syndrome—sphingosine-1-phosphate lyase (SGPL1) deficiency: 6 cases and a review of the literature. Pediatr Nephrol. 2023;38:711–9, http://dx.doi.org/10.1007/s00467-022-05656-5.
- Dorval G, Kuzmuk V, Gribouval O, Welsh GI, Bierzynska A, Schmitt A, et al. TBC1D8B loss-of-function mutations lead to X-linked nephrotic syndrome via defective trafficking pathways. Am J Hum Genet. 2019;104:348–55, http://dx.doi.org/10.1016/j.ajhg.2018.12.016.
- Fang Z, Zhang C, Jin Y, Tong J, Liu J, Hao X, et al. Adult-onset focal segmental glomerulosclerosis with steroid-dependent nephrotic syndrome caused by a novel TBC1D8B variant: a case report and literature review. Am J Kidney Dis. 2023;81:240–4, http://dx.doi.org/10.1053/j.ajkd.2022.06.012.
- Bullich G, Vargas I, Trujillano D, Mendizábal S, Piñero-Fernández JA, Fraga G, et al. Contribution of the TTC21B gene to glomerular and cystic kidney diseases. Nephrol Dial Transplant. 2017;32:151–453156, http://dx.doi.org/10.1093/ndt/gfv453.
- Cong EH, Bizet AA, Boyer O, Woerner S, Gribouval O, Filhol E, et al. A homozygous missense mutation in the ciliary gene TTC21B causes familial FSGS. J Am Soc Nephrol. 2014;25:2435–43, http://dx.doi.org/10.1681/ASN.2013101126.
- Harshman LA, Ng BG, Freeze HH, Trapane P, Dolezal A, Brophy PD, et al. Congenital nephrotic syndrome in an infant with ALG1-congenital disorder of glycosylation. Pediatr Int. 2016;58:785–8, http://dx.doi.org/10.1111/ped.12988.
- Sadowski CE, Lovric S, Ashraf S, Pabst WL, Gee HY, Kohl S, et al. A single-gene cause in 29.5% of cases of steroid-resistant nephrotic syndrome. J Am Soc Nephrol. 2015;26:1279–89, http://dx.doi.org/10.1681/ASN.2014050489.
- 92. Lipska-Zietkiewicz BS. Genetic Steroid-Resistant Nephrotic Syndrome Overview 1. Clinical Characteristics of Genetic Steroid-Resistant Nephrotic Syndrome 2. Causes of Genetic

Steroid-Resistant Nephrotic Syndrome. GeneReviews InternetReviews. Published online 2022:1-15 [Accessed 15 June 2024]. Available from:

https://www.ncbi.nlm.nih.gov/books/NBK573219/.

- Boyer O, Mollet G, Dorval G. Neurological involvement in monogenic podocytopathies. Pediatr Nephrol. 2021;36:3571–83, http://dx.doi.org/10.1007/s00467-020-04903-x.
- 94. Park E, Ahn YH, Kang HG, Yoo KH, Won NH, Lee KB, et al. COQ6 mutations in children with steroid-resistant focal segmental glomerulosclerosis and sensorineural hearing loss. Am J Kidney Dis. 2017;70:139–44, http://dx.doi.org/10.1053/j.ajkd.2016.10.040.
- 95. Adutwum M, Hurst A, Mirzaa G, Kushner JD, Rogers C, Khalek N, et al. Six new cases of CRB2-related syndrome and a review of clinical findings in 28 reported patients. Clin Genet. 2023;103:97–102, http://dx.doi.org/10.1111/cge.14222.
- 96. Yang J, Xu Y, Deng L, Zhou L, Qiu L, Zhang Y, et al. CUBN gene mutations may cause focal segmental glomerulosclerosis (FSGS) in children. BMC Nephrol. 2022;23:1–8, http://dx.doi.org/10.1186/s12882-021-02654-x.
- 97. Silva CAB, Moura-Neto JA, Dos Reis MA, Vieira Neto OM, Barreto FC. Renal manifestations of fabry disease: a narrative review. Can J Kidney Health Dis. 2021;8, http://dx.doi.org/10.1177/2054358120985627.
- 98. Boyer O, Nevo F, Plaisier E, Funalot B, Gribouval O, Benoit G. INF2 mutations in Charcot–Marie–Tooth disease with glomerulopathy. N Engl J Med. 2011;365:2377–88, http://dx.doi.org/10.1056/nejmoa1109122.
- 99. Rood IM, Deegens JKJ, Wetzels JFM. Genetic causes of focal segmental glomerulosclerosis: implications for clinical practice. Nephrol Dial Transplant. 2012;27:882–90, http://dx.doi.org/10.1093/ndt/gfr771.
- 100. AbuMaziad AS, Abusaleh R, Bhati S. Congenital nephrotic syndrome. J Perinatol. 2022;41:2704–12, http://dx.doi.org/10.1038/s41372-021-01279-0.
- 101. Boyer O, Woerner S, Yang F, Oakeley EJ, Linghu B, Gribouval O, et al. LMX1B mutations cause hereditary FSGS without extrarenal involvement. J Am Soc Nephrol. 2013;24:1216–22, http://dx.doi.org/10.1681/ASN.2013020171/-/DCSUPPLEMENTAL.
- 102. Kopp JB. An expanding universe of FSGS genes and phenotypes: LMX1B mutations cause familial autosomal dominant FSGS lacking extrarenal manifestations. J Am Soc Nephrol. 2013;24:1183–5, http://dx.doi.org/10.1681/ASN.2013060661.
- 103. Lovric S, Ashraf S, Tan W, Hildebrandt F. Genetic testing in steroid-resistant nephrotic syndrome: when and how? Nephrol Dial Transplant. 2016;31:1802–13, http://dx.doi.org/10.1093/ndt/gfv355.
- 104. Singh N, Nainani N, Arora P, Venuto RC. CKD in MYH9-related disorders. Am J Kidney Dis. 2009;54:732–40, http://dx.doi.org/10.1053/j.ajkd.2009.06.023.

- 105. Barua M, Stellacci E, Stella L, Weins A, Genovese G, Muto V, et al. Mutations in PAX2 associate with adult-onset FSGS. J Am Soc Nephrol. 2014;25:1942–53, http://dx.doi.org/10.1681/ASN.2013070686.
- 106. Hu R, Wen Y, Ye W, Zhang L, Si N, Zheng K. FSGS in Chinese twins with a de novo PAX2 mutation: a case report and review of the literature. J Nephrol. 2021;34:2155–8, http://dx.doi.org/10.1007/S40620-021-01055-5/METRICS.
- 107. Vivante A, Chacham OS, Shril S, Schreiber R, Mane SM, Pode-Shakked B, et al. Dominant PAX2 mutations may cause steroid-resistant nephrotic syndrome and FSGS in children. Pediatr Nephrol. 2019;34:1607–13, http://dx.doi.org/10.1007/s00467-019-04256-0.
- 108. López LC, Schuelke M, Quinzii CM, Kanki T, Rodenburg RJ, Naini A, et al. Leigh syndrome with nephropathy and CoQ10 deficiency due to decaprenyl diphosphote synthase subunit 2 (PDSS2) mutations. Am J Hum Genet. 2006;79:1125–9, http://dx.doi.org/10.1086/510023.
- 109. Lovric S, Goncalves S, Gee HY, Oskouian B, Srinivas H, Choi WI, et al. Mutations in sphingosine-1-phosphate lyase cause nephrosis with ichthyosis and adrenal insufficiency. J Clin Invest. 2017;127:912–28, http://dx.doi.org/10.1172/JCI89626.
- 110. Prasad R, Hadjidemetriou I, Maharaj A, Meimaridou E, Buonocore F, Saleem M, et al. Sphingosine-1-phosphate lyase mutations cause primary adrenal insufficiency and steroid-resistant nephrotic syndrome. J Clin Invest. 2017;127:942–53, http://dx.doi.org/10.1172/JCI90171.
- 111. Ahn YH, Park EJ, Kang HG, Kim SH, Cho HY, Shin JI, et al. Genotype–phenotype analysis of pediatric patients with WT1 glomerulopathy. Pediatr Nephrol. 2017;32:81–9, http://dx.doi.org/10.1007/s00467-016-3395-4.
- 112. Chernin G, Vega-Warner V, Schoeb DS, Heeringa SF, Ovunc B, Saisawat P, et al. Genotype/phenotype correlation in nephrotic syndrome caused by WT1 mutations. Clin J Am Soc Nephrol. 2010;5:1655–62, http://dx.doi.org/10.2215/CJN.09351209.
- 113. Agarwal AK, Zhou XJ, Hall RK, Nicholls K, Bankier A, Van Esch H, et al. Focal segmental glomerulosclerosis in patients with mandibuloacral dysplasia owing to ZMPSTE24 deficiency. J Investig Med. 2006;54:208–13, http://dx.doi.org/10.2310/6650.2006.05068.
- 114. Hermle T, Schneider R, Schapiro D, Braun DA, van der Ven AT, Warejko JK, et al. GAPVD1 and ANKFY1 mutations implicate RAB5 regulation in nephrotic syndrome. J Am Soc Nephrol. 2018;29:2123–38, http://dx.doi.org/10.1681/ASN.2017121312.